

**IN THE SPECIFICATION:**

**Please amend the paragraph beginning on page 3, line 24 as follows:**

In a preferred embodiment, the knock-in vectors of the present invention include a nucleotide sequence coding for an  $\alpha$ -1,2-mannosidase or a functional part thereof and are capable of expressing the  $\alpha$ -1,2-mannosidase or the functional part in a methylotrophic yeast strain. A preferred nucleotide sequence is a nucleotide sequence encoding the  $\alpha$ -1,2-mannosidase of a fungal species, and more preferably, *Trichoderma reesei*. Preferably, the  $\alpha$ -1,2-mannosidase expression vector is engineered such that the  $\alpha$ -1,2-mannosidase or a functional part thereof expressed from the vector includes an ER-retention signal. A preferred ER-retention signal is HDEL (SEQ ID NO: 1). The  $\alpha$ -1,2-mannosidase coding sequence can be operable linked to a constitutive or inducible promoter, and a 3' termination sequence. The vectors can be integrative vectors or replicative vectors. Particularly preferred  $\alpha$ -1,2-mannosidase expression vectors include pGAPZMFManHDEL, pGAPZMFManMycHDEL, pPICZBMFManMycHDEL, pGAPZmManHDEL, pGAPZmMycManHDEL, pPIC9mMycManHDEL and pGAPZmMycManHDEL.

**Please amend the paragraph beginning on page 4, line 10 as follows:**

In another preferred embodiment, the knock-in vectors of the present invention include a sequence coding for a glucosidase II or a functional part thereof and are capable of expressing the glucosidase II or the functional part in a methylotrophic yeast strain. A preferred nucleotide sequence is a nucleotide sequence encoding the glucosidase II of a fungal species, and more preferably, *Saccharomyces cerevisiae*. Preferably, the glucosidase II expression vector is engineered such that the glucosidase II or a functional part thereof expressed from the vector

includes an ER-retention signal. A preferred ER-retention signal is HDEL (SEQ ID NO: 1). The glucosidase II coding sequence can be operable linked to a constitutive or inducible promoter, and a 3' termination sequence. The vectors can be integrative vectors or replicative vectors. Particularly preferred glucosidase II expression vectors include pGAPZAGLSII, pPICZAGLSII, pAOX2ZAGLSII, pYPTIZAGLSII, pGAPADEglsII, pPICADEglsII, pAOX2ADEglsII, pYPTIADEglsII, pGAPZAglsIIHDEL and pGAPADEglsIIHDEL.

**Please amend the paragraph beginning on page 40, line 5 as follows:**

The cloning of a *Trypanosoma cruzi* *trans*-sialidase gene coding for an active *trans*-sialidase member without the C-terminal repeat domain has been described by Laroy et al. (*Protein Expression and Purification* 20: 389, 2000) which is incorporated herein by reference. The sequence of this *Trypanosoma cruzi* *trans*-sialidase gene is available through NCBI Genbank under the Accession No. AJ276679. For expression in *P. pastoris*, the entire gene was cloned in pHILD2 (Invitrogen, San Diego, CA), creating pHILD2-TS. To allow better secretion, pPIC9-TS was created in which *trans*-sialidase was linked to the prepro secretion signal of the yeast  $\alpha$ -mating factor. Plasmids pPIC9-TSE and pCAGGS-prepro-TSE were created where the epitope E-tag was added to the C-terminal of the *trans*-sialidase to allow easy detection and purification. The construction of pHILD2-TS, pPIC9-TSE and pCAGGS-prepro-TSE has been described by Laroy et al. (2000), incorporated herein by reference. The vectors used in the construction were made available through <http://www.belspo.be/becm/lmbp.htm#main> for pCAGGS (No. LMBP 2453), Invitrogen, San Diego, CA for pHILD2 and pPIC9, and Pharmacia Biotech for pCANTAB-5E.

**Please amend the paragraph beginning on page 44, line 9 as follows:**

First, the full ORF of the *Pichia pastoris* Och1 gene was PCR cloned in pUC18 to obtain plasmid pUC18pOch1. pUC18pOch1 was cut with HindIII, blunt-ended with T4 polymerase, then cut with XbaI, releasing a fragment containing the 5' part of the *Pichia pastoris* Och1 gene. This fragment was ligated into the vector pBLURA IX (available from the Keck Graduate Institute, Dr. James Cregg, <http://www.kgi.edu/html/noncore/faculty/cregg/cregg.htm>), which had been cut with *Eco* RI, blunt-ended with T4 polymerase, and then cut with *Nhe* I. This ligation generated pBLURA5'PpPCH1, as shown in **Figure 8**.